

Original Research Article

<http://dx.doi.org/10.20546/ijcmas.2016.507.044>

Evaluation of the Efficacy of the Compound Actinomycin D in Tumor Cell Lines

R. Cibi and A. Jayakumaran Nair*

Department of Biotechnology, University of Kerala, Karyavattom, Trivandrum, India

*Corresponding author

ABSTRACT

Keywords

Actinomycin D,
Cytotoxicity,
Acridine orange
and ethidium
bromide staining,
Comet assay.

Article Info

Accepted:

15 June 2016

Available Online:

10 July 2016

The search for anticancer drug has lead researchers to study actinomycin D, a chromopeptide antibiotic that destroy cancer cells. The antitumor activity of actinomycin D analyzed on different cell lines by MTT assay. Different concentrations of compound (0.01-1.25 μ g/mL) was added to cell lines and incubated for 72 hrs. The compound actinomycin D showed cytotoxicity against U251, HCT-116 and MCF-7 cancer cell lines depending on the dose and incubation time. The best activity was shown against U251 cell line with IC₅₀ value of 1.07 μ g/ml on 24 hr, 0.56 μ g/ml on 48 hr and 0.028 μ g/ml on 72hr. The morphological change in actinomycin D treated U251 cells were detected by acridine orange and ethidium bromide staining. The comet assay revealed that actinomycin D inhibited the growth of U251 cells by induction of DNA damage.

Introduction

Actinomycin D is the best known compound from the actinomycin group whose wide application potential has been extensively studied. Actinomycin D is a orange red compound having two cyclic pentapeptide lactones attached to 2 amino-4,6-dimethyl-3-oxo-phenoxazine -1,9-dicarboxylic acid, first reported in *Streptomyces antibioticus* (Waksman and Woodruff, 1940). Subsequently number of other bacteria were also reported to produce actinomycin D including *Streptomyces fradiae* (Bossi *et al.*, 1958), *Streptomyces parvulus* (Williams and Katz, 1977), *Streptomyces plicatus* (Lam *et al.*, 2002), *Streptomyces violascens* (Roos and Loane, 2004), *Streptomyces sindenensis*

(Praveen *et al.*, 2008) etc. Actinomycin D is found to be the most effective natural analog for the treatment of cancer in children and adults. It has been used clinically as a chemotherapeutic agent for the treatment of Wilm's tumor with 90% cure rate (Green, 1997; Farber *et al.*, 2002) and rhabdomyosarcoma (Womer, 1997). This compound was approved for the treatment of pediatric solid tumors by US FDA in 1964 (Newman and Cragg, 2007). The biological activity is due to the ability of actinomycin D to intercalate with the DNA double helix and there by inhibiting DNA directed RNA synthesis, this prevents unwinding of the DNA to facilitate its

interaction with RNA polymerase (Chou *et al.*, 2002; Bendic, 2007).

The exciting potentialities of actinomycin D have attracted researchers to explore the medicinal value of actinomycin D. The present study was to investigate the cytotoxic potency against different tumor cell lines and quantitative detection of DNA damage induced by compound.

Materials and Methods

Actinomycin D

The compound actinomycin D used in the present work was extracted from *Streptomyces parvulus* using methanol and purified by TLC following a standard protocol (Shetty, 2014). The compound was characterized using JEOL JMS 600 H mass spectrometer (JOEL, USA).

Cytotoxicity assay

The cytotoxicity activity of actinomycin D against human tumor cell lines HCT-116 (colon adenocarcinoma), MCF-7 (breast carcinoma) U251 (glioblastoma) was determined by MTT (3-[4, 5-dimethylthiazol-2-yl]-2, 5-diphenyl tetrazolium bromide) assay (Wilson, 2000). Briefly 1×10^4 cells per well were seeded at log phase in 96 well plates. After cultivation for 24hrs the media was replaced with fresh media containing different concentration (0.01-1.25 $\mu\text{g/ml}$) of actinomycin D dissolved in 0.1% DMSO and treated for 24, 48 and 72 hr. After incubation at 37°C in 5% CO_2 , 100 μl of medium containing 20 μl of MTT solution (5mg/ml in PBS) was added to each well and cells were incubated for 4hrs at 37°C . At the end incubation, medium was removed and 100 μl of DMSO was added to each well for formazan crystals to dissolve. The absorbance was

measured at 570nm using Mullikan GO Micro plate Spectrophotometer (Thermoscientific). Doxorubicin was used for the positive control. The growth inhibition was calculated from the formulae $(1-A/B) \times 100\%$ where A and B corresponds to the mean absorbance of treated and control wells. The concentration of the compound that gives 50% inhibition was expressed as the IC_{50} ($\mu\text{g/ml}$).

Acridine orange and Ethidium bromide staining

To detect morphological changes induced by actinomycin D, briefly 1×10^4 U251 cells per well were seeded at log phase in 96 well plates and treated with 0.028 $\mu\text{g/ml}$ of actinomycin D for 72 hr. The untreated and treated cells were stained 20 μl of acridine orange and ethidium bromide (100 $\mu\text{g/ml}$ each) and observed under a fluorescence microscopy (Leica, Germany).

Comet assay

Measurement of cellular DNA damage induced by compound was carried out using alkaline comet assay (Collins *et al.*, 2008; Ramos *et al.*, 2010). Briefly, U251 cells were seeded in 24 well plates at a density of 1×10^5 cells per well for 24hr at 37°C . After adhesion cells were treated with 0.028 $\mu\text{g/ml}$ and 1.25 $\mu\text{g/ml}$ of actinomycin D and incubated for 72h. After treatment, cells in each well were trypsinized, centrifuged and resuspended in 1ml PBS. Samples were stored at 4°C . A 10 μl aliquot of resuspended cells were mixed with 70 μl of 0.5% (w/v) low melting point agarose in PBS and transferred to 1% (w/v) agarose coated slides (normal melting point). Slides were immersed in lysis buffer (10mM Tris, 2.5M NaCl, 100mM EDTA, pH adjusted to 10 before addition of 1% Triton X-100) and kept at 4°C . After lysis the slides were

immersed in an alkaline electrophoresis buffer (300mM NaOH, 1Mm EDTA, pH 13) for 30 min at 4°C. After a 20 min of electrophoresis by applying a current of 250mA, slides were removed and neutralized with two 5-min washes in a solution of 0.4 M Tris –HCl (pH 7.5), then fixed with 100% ethanol and dried at room temperature. After that slides were stained with ethidium bromide (20µg/ml) and examined under fluorescence microscope DNA migration was determined based on tail intensity (% DNA in tail) using a casplab software.

Result and Discussion

Cytotoxicity assay

To determine the cytotoxic activity of actinomycin D to tumor cell lines the cells were treated with increasing concentration of actinomycin D. The MTT cytotoxicity assay revealed that actinomycin D decreased the cell viability significantly (P<0.05) in a concentration dependent manner. As shown in Fig.1 U251 cells were more sensitive to actinomycin D compared to HCT and MCF7. Table 1 shows the IC50 values of actinomycin D and doxorubicin against U251, HCT and MCF7. Actinomycin D showed a cytotoxic effect at an IC50 of 0.9µg/ml to U251 cells and to 1.09µg/ml to HCT-116 after 24hrs of treatment. However

no IC50 was found on MCF7 at the same treatment duration. At 48hr the actinomycin D exhibited lowest IC50 Of 0.56µg/ml to U251 cells in contrast with the IC50 value of 1.03µg/ml to Hct-116 cells and IC50 value of 0.9µg/ml to MCF7 cells. When the treatment was prolonged to 72 hr, actinomycin D exhibited IC50 value of 0.028µg/ml to U251, IC50 value of 0.55µg/ml to HCT-116 and 0.09µg/ml against MCF7. The IC50 of purified actinomycin D comparable to standard doxorubicin.

There are reports of chemotherapeutic agents capable of inducing apoptosis, inhibiting cell proliferation or modulating signal transduction for the treatment of cancer (Bakshi *et al.*, 2010). The results agree with previous investigation that actinomycin D has been shown to have cytotoxic activity against pancreatic cancer cell lines by inducing apoptosis (Kleeff *et al.*, 2000).

Acridine orange and Ethidium bromide staining

The morphological changes suggestive of apoptosis were observed in U251 cells when treated with 0.028µg/ml actinomycin D after 72 hrs was confirmed when cells were viewed after staining under fluorescence microscope Fig. 2.

Table.1 IC50 values of actinomycin D and doxorubicin against different tumor cell lines

Compound	U251			Hct-116			MCF7		
	24hrs	48hrs	72hrs	24hrs	48hrs	72hrs	24hrs	48hrs	72hrs
Actinomycin D	0.9 µg/ml	0.56 µg/ml	0.028 µg/ml	1.09 µg/ml	1.03 µg/ml	0.55 µg/ml	-	0.9 µg/ml	0.09 µg/ml
Doxorubicin	-	1.15 µg/ml	0.5 µg/ml	0.9 µg/ml	0.48 µg/ml	0.18 µg/ml	1.20 µg/ml	1.1 µg/ml	0.84 µg/ml

Table.2 DNA damage induced by actinomycin D in U251 cell line

Parameter	Untreated	Treated(0.02µg/ml)	Treated (1.25µg/ml)
Tail length	6	17.33	26.93
Tail moment	0.01	2.11	11.65
% DNA in tail	99.79	87.59	44.594
OTM	0.1174	4.41	10.58

Fig. 1 Effect of actinomycin D (0.01 – 1.2µg/ml) on different tumor cell lines U251, HCT-116 and MCF7

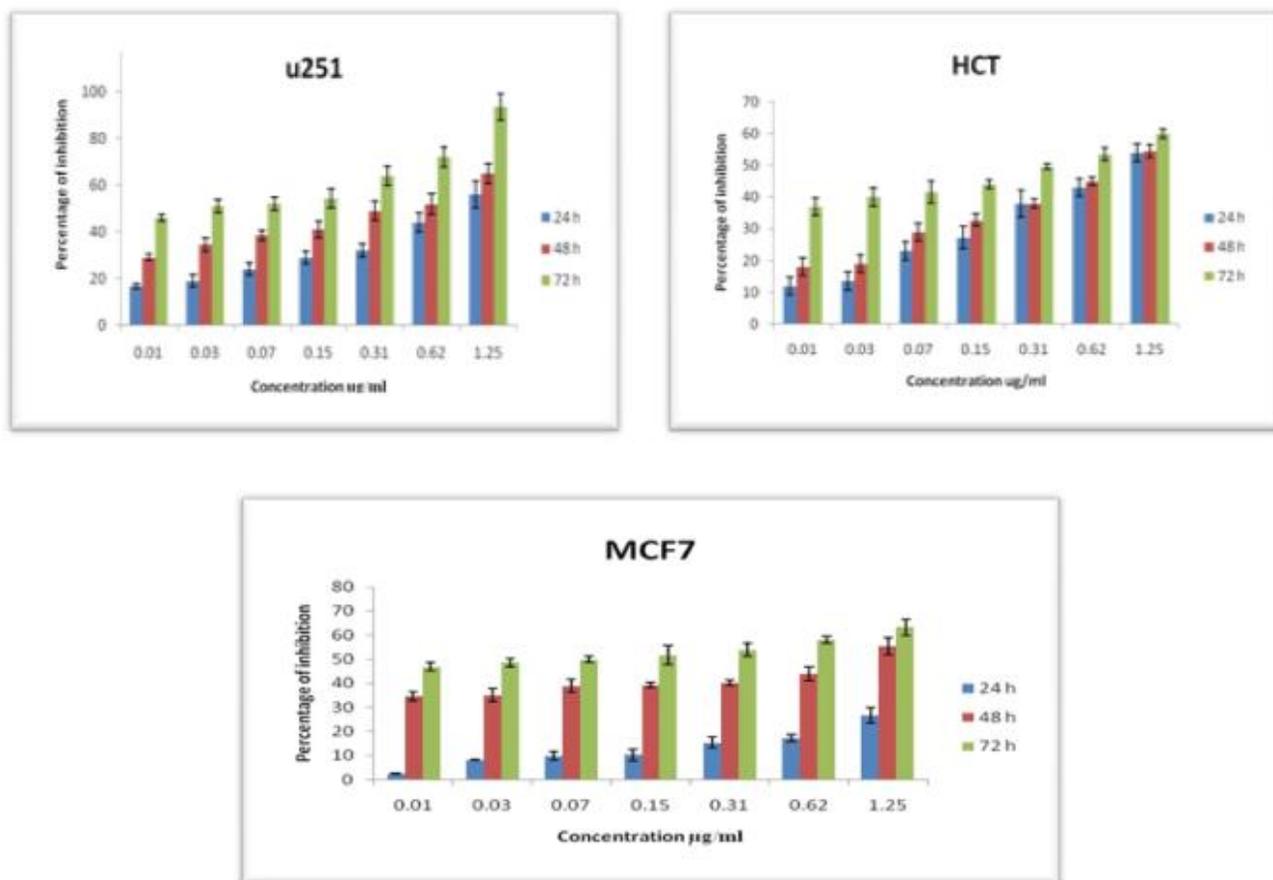
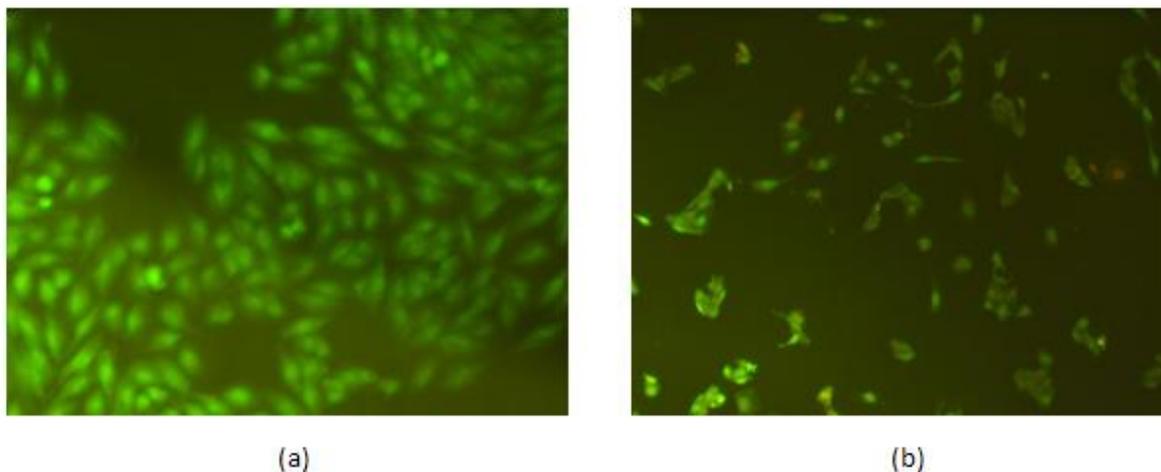


Fig. 2 Fluorescent microscopic images of actinomycin D treated U251 cell lines (a) control (b) treated



The morphological changes induced by actinomycin D in U251 cells was further confirmed by acridine orange and ethidium bromide staining. There are reports of commercially available actinomycin D (0.05 μ g/ml) inhibiting the growth of 60% glioblastoma cells after 92 hr by activating caspase-3 and 4 followed by apoptotic cell death (Narita *et al.*, 2000). The concentration of compound increased the rate of inhibition in a dose dependant manner. Previous reports have shown that low doses of actinomycin D inhibit the cell proliferation of neuroblastoma cell lines by inducing apoptosis(Cortes *et al.*, 2016).

Comet assay

Actinomycin D induce DNA strand breakage and formation of comets in U251 cells and in the untreated cells the nuclei were circular without formation of any visible tail as shown in Fig. 3. Percentage of cells with tail was significantly higher (P value ≤ 0.05) than the untreated cells. Tail lengths of the comets in cells treated with 0.028 μ g/ml was (17.33) and in 1.25 μ g/ml was found to be (26.93) and tail moment in 0.028 μ g/ml treated cell was 2.11 and in cells

treated with 1.25 μ g/ml found to be 11.65 shown in Table 2.

The level of DNA damage induced by compound to U251 cells was quantified by determining the intensity of DNA tail by comet assay(Tice *et al.*, 2000, Hartmann *et al.*, 2003)The present observation from comet assay indicates that cytotoxic effect induced by actinomycin D towards U251 may be caused by DNA damage.

In conclusion, the study has demonstrated for testing the cytotoxicity activity of actinomycin D against U251, Hct-116 and MCF-7cancer cell line. The U251 cells employed to assess the DNA damage induced by actinomycin D. The present study confirmed that actinomycin D induced potent cytotoxic activity and genotoxicity in U251 cells.

Acknowledgement

We are thankful to the University of Kerala for the research fellowship and infrastructure provided by Department of Biotechnology, University of Kerala is also acknowledged.

References

- Bakshi, H., Sam, S., Rozati, R., Sultan, P., Islam, T., Rathore, B., Lone, Z., Sharma, M., Tripathi, J., and Saxena, R. 2010. DNA Fragmentation and Cell Cycle Arrest: A Hallmark of apoptosis induced by crocin from Kashmiri saffron in a human pancreatic cancer cell line. *Asian Pac J. Cancer Prev.*, 11: 675-679.
- Bentic, C., Enache, M., and Volanschi, E. 2005. Analysis of actinomycin D - DNA model complexes using a quantum chemical criterion. *J. Mol. Graph. Model*, 24: 10-16.
- Bossi, J., Hutter, R., Schierlein, K.W., Neipp, L., and Zaheer, H. 1958. Metabolic products from actinomycetes XIV. Actinomycin Z. *HelvchimActa*, 41: 1645-1652.
- Chou, S.H., Chin, K.H., and Chen, F.M. 2002. Looped out and perpendicular : Deformation of Watson -Crick base pair associated with actinomycin D binding. *Proc. Natl. Acad. Sci.*, 99: 6625-6630.
- Collins, A.R., Oscoz, A.A., Brunborg, G., Gaivão, I., Giovannelli, L., Kruszewski, M., et al. 2008. The comet assay: Topical issues. *Mutagenesis*, 23: 143-51.
- Cortes, C.L., Veiga, S.R., Almacellas, E., Losa, H. J., Ferreres, J.C, Kozma, S.C., Ambrosio, S., Thomas, G., and Tauler, T. 2016. Effect of low doses of actinomycin D on neuroblastoma cell lines. *Mol. Cancer*, 15: 1.
- Farber, S., D'Angio, G., Evans, A., and Mitus, A. 2002. Clinical studies of actinomycin D with special reference to Wilms' tumor in children. *J. Urol.*, 168: 2560 - 2562.
- Green, D.M. 1997. Paediatric oncology update/Wilm' tumour. *Eur. J. Cancer.*, 33: 409-418.
- Hartmann, A., Agurell, E., Beevers, C., Brendler-Schwaab, S., Burlinson B., Clay, P et al.,. 2003. Recommendation for conducting that invivo alkaline comet assay. In. 4th international comet assay workshop. *Mutagenesis*. 18: 45-51.
- Kleeff, et al. 2000. Actinomycin D induces apoptosis and inhibits growth of pancreatic cancer cells. *Int. J. Cancer.*, 86: 399.
- Lam, K.S., Gustavson, D.R., Pirnik, D.L., Pack, E., Bulanhagui, C., Mamber, S.W., Forenza, S., Stodieck, L.S., and Klaus, D.M. 2002. The effect of space flight on the production of actinomycin D by *Streptomyces plicatus*. *J. Ind. Microbiol. Biotechnol.*, 29: 299-302.
- Narita, Y., Asai, A., Kuchino, Y., and Kirino, T. 2000. Actinomycin D and staurosporine, potent apoptosis inducers in vitro, are potentially effective chemotherapeutic agents against glioblastoma multiforme. *Cancer. Chemother. Pharmacol.*, 45: 149-156.
- Newman, D.J. and Cragg, G.M.J. 2007. Natural products as sources of new drugs over the last 25 years. *J. Nat. Prod.*, 70: 461-477.
- Praveen, V., Tripathi, C.K.M, Bihari, V., and Srivastava, S.C. 2008b. Production of actinomycin-D by the mutant of a new isolate of *Streptomyces sindenensis*. *Braz. J. Microbiol.*, 39: 689-692.
- Ramos, A.A., Azqueta, A., Pereira-Wilson, C., Collins, A.R. 2010. Polyphenolic compounds from *Salvia* species protect cellular DNA from oxidation and stimulate DNA repair in cultured human cells. *J. Agric. Food Chem.*, 58: 7465-71.
- Roos, G.H.P. and Loane, C. 2004. A clarifying NMR study of actinomycin

- X2. *J. Saudi. Chem. Soc.*, 8: 289-294.
- Shetty, P.S., Buddana, K.S., Tatipamula, V.B., Naga, Y.V. and Ahmad, J. 2014. Production of polypeptide antibiotic from *Streptomyces parvulus* and its antibacterial activity. *Braz J Microbiol.*, 45: 301-312.
- Tice, R.R., Agurell, E., Anderson, D., Burlinson, B., Hartmann, A., Kobayashi H *et al.* 2000 Single cell gel/ comet assay: guidelines for in vitro and in vivo genetic toxicology testing. *Environ. Mol. Mutagen.*, 35: 206-21.
- Waksman, S. and Woodruff, H.B. 1940. Bacteriostatic and bactericidal substances produced by a soil Actinomycetes. *Proc. Soc. Exp. Biol. Med.*, 45: 609-614.
- Williams, W.K. and Katz, E. 1977. Development of chemically defined medium for the synthesis of actinomycin D by *Streptomyces parvulus*. *Antimicrob. Agents. Chemother.*, 11: 281-290.
- Wilson, A.P. 2000. Cytotoxicity and viability assays in animal cell culture. A practical approach (3rd edn). Oxford Univ Press. Oxford PP, 175-21.
- Womer, R.B. 1997. Soft tissue sarcomas. *Eur. J. Cancer*, 33: 2230 –2234.

How to cite this article:

Cibi, R., and Jayakumaran Nair, A. 2016. Evaluation of the Efficacy of the Compound Actinomycin D in Tumor Cell Lines. *Int.J.Curr.Microbiol.App.Sci.* 5(7): 405-411.
doi: <http://dx.doi.org/10.20546/ijcmas.2016.507.044>